

The Role of Some Cytokines in women with Recurrent Abortion in Iraqi Women

ROUA JAMAL ABDULKHALIQ¹, SABAATAHER MOHAMMED², AHMED ABDUL-HASSAN ABBAS³

ABSTRACT

Recurrent spontaneous abortion (RSA) in the general populations is very common occurrence and often changes among different communities. This study was designed to measure some cytokines in the case of recurrent spontaneous abortion and healthy pregnant. Blood samples were collected from 210 women (180 women with recurrent abortion (three or more abortions) and 30 women with normal pregnancy to three or more birth and without a previous abortion and without any infection) in the first trimester. Based on clinical examination and diagnostic laboratory findings of rapid test and ELISA for TORCH. Measured levels of immunoglobulin (IgM and IgG) for (*Toxoplasmosis, cytomegalovirus, herpes and rubella*) in normal pregnant and aborted women. Depending on the results of tests, the patients were divided into four groups: group one included 30 women with recurrent abortion with sero-negative for TORCH test was (17%), group two recurrent aborted with sero-positive for anti-toxoplasma antibodies was (24.8%), group three included women with a healthy pregnancy promise as a control group was (14.3%) while group four recurrent abortion suffering from infected with different causes such as (*cytomegalovirus, Rubella or Herpes*) ratio was (43.8%). Then taking thirty serum samples of women from the first and second group with recurrent abortion and compared with control group then measured serum levels of pro-inflammatory cytokines Interleukin-6 (IL-6) and anti-inflammatory marker Tumor growth factor-beta1 (TGF- β 1). The results showed that the serum levels of IL-6 was highest in group 1 and 2 (474.50 pg/ml and 629.60 pg/ml) respectively compared with control group (247.03 pg/ml), there was a significant difference. While TGF- β 1 was highest level in group 1 (2176.30 pg/ml) followed by group 2 (1710.63 pg/ml) then control group (1099.76 pg/ml) with significant difference between three groups. The results indicated that the serum levels of pro-inflammatory and anti-inflammatory marker found with high level in recurrent abortion compared with healthy pregnant.

Keywords: Abortion, cytokines, pregnancy

INTRODUCTION

Spontaneous abortion (SA) is the most common complication of early pregnancy, refers to one of the most frequent reproductive events is recurrent spontaneous abortion (RSA). It is defined as three or more repeated pregnancy losses before the fetus has reached a viable gestational age¹. Reasonably accepted etiologic causes include, genetics, immunologic factors, placental abnormality, endocrine disorder, nutritional, environmental factors and infection with microorganisms like *Toxoplasma gondii*, *Cytomegalovirus*, *siphilis*, *rubella*, *herpes* and maternal disease (such as diabetes mellitus, thyroid disease)². Toxoplasmosis caused abortions usually occur during the first half of gestation and effects on liver and spleen functions³. When the congenital toxoplasmosis occurs early in pregnancy, it may lead to severe damage or abortion. Embryo acts as an

allograft to the mother's body, it is remaining normally in the mother's womb during the entire gestational period in the case of normal successful pregnancy⁴. The placental barrier is traffic of cytotoxic cells to the fetus and cytotoxic antibodies are removed by the placenta before they reach the fetal circulation⁵. T cells may play an important role in pre-implantation and embryo development in implantation process and in the phenomenon of fetal allograft tolerance⁶. Th₁ cytokine inhibit Th₂ cells expansion and Th₂ cytokines block activation of Th₁ cells⁷. IL-6 might have both beneficial effects and detrimental effects on the events of early pregnancy also implicated in the pathophysiology of abnormal pregnancies and other disease such as Rheumatoid Arthritis, Autoimmune disease, preeclampsia and obesity^{8,9,10}. TGF- β in the presence of IL-6 can promote inflammation and autoimmune conditions¹¹.

The aims of the present study were to detect the serum levels of anti-inflammatory cytokines (TGF- β) and pro-inflammatory cytokines (IL-6) in women with recurrent abortion.

MATERIAL AND METHODS

¹College of Veterinary medicine University of Fallujah.

²College of Science AL-Mustansiriya University.

³College of Medicine, Al-Nahrain University.

Correspondence to Dr. Roua Jamal Abdul Khaliq Email: roua.jamal@gmail.com

Patients and control: The current study included 180 women with recurrent abortion during the first-trimester and 30 women as a control groups were with normal third delivery or more and with no previously recognized miscarriage. The ages of these women were ranged between 20-35 years. The total number of women (210) was referred to Obstetrics and Gynecology Department of AL-Yarmook Teaching Hospital, Baghdad Teaching Hospital and AL-Batool Teaching Hospital during the period from March to December 2016.

Samples collection: The 5 ml of venous blood was taken at the time of miscarriage by using sterile disposable syringes, (3ml) of blood placed in a plain tube with gel clot and left to stand for one hour at room temperature for clot formation, for serum collection, the tube centrifuged for 10 minutes at 3000 rpm. Then the serum aspirated by using a Pasteur pipette and dispensed into sterile eppendorf tube and stored at -20 C until used.

TORCH rapid Test for IgM and IgG: Practical work was done according to the instructions of manufacturers (Ecotest, China).

Enzyme Linked Immunosorbent Assay for the detection of IgG or IgM antibodies in human serum: The practical work was done according to the instructions of manufacturers (Diagnostic Automation, INC, USA).

Study groups: According to TORCH and ELISA test divided into:

-Group one: women with repeated aborted three or more during the first trimester with recurrent abortion with sero-negative for TORCH test

-Group two: women with repeated aborted three or more during the first trimester with sero-positive for anti-toxoplasma antibodies.

-Group three: women with normal third delivery or more and with no previously recognized miscarriage.

-Group four: women with repeated aborted three or more during the first trimester with different causes.

Selected thirty samples were taken from each three groups for determined levels of cytokines.

Measurement of cytokines:

Human IL-6 and TGF-β1 ELISA Kits: The practical work was done according to the instructions of manufacturers (Komabiotechinc / korea).

Statistical analysis: Statistical analysis was performed with SPSS22. Numerical data were

described as mean, standard deviation and least significant difference (LSD) test was used to significant compare between groups. Results were analyzed by comparison between groups by one way ANOVA.

RESULTS

Table 1 shows the distribution of studied groups according-to TORCH results by lateral flow chromatographic immunoassay and ELISA TORCH as following divided into group1: recurrent abortion with Unknown causes the percentage was 36(17.1%), group2: recurrent abortion with Toxoplasma only was 52(24.8%), group3: healthy women with normal pregnancy (control group) was 30 (14.3%) and group 4: recurrent abortion with different causes was 92(43.8%).

Seroprevalence of IgM and IgG among study groups: Table (2) Illustrates seroprevalence of IgM and IgG antibody for pathological causes among study groups according-to ELISA test. The seropositive Antibodies of IgM and IgG for *Toxoplasma gondii* were 18(8.5%), 30(14.3%) followed by *Cytomegalovirus* were 17(8.09%), 28 (13.3%), *Rubella* were 9(4.3%), 15(7.1%), HSV were 3(1.5%), 8(3.8%), while for healthy pregnant were 0(0%), 0(0%) respectively. The seroprevalence of (IgM and IgG) antibody for *Toxoplasma gondii* was 4(1.9%), *Cytomegalovirus* was 5(2.4%), *Rubella* was 6(2.9%), *Herpes Simplex Virus* was 1(0.4%) and Healthy pregnant was 0 (0%).

The mean of IL-6 in the group1 was (474.50±158.86) pg/ml, group 2 was (629.60±115.45) pg/ml and in group 3 was (247.03±46.00) pg/ml as shows in the table 3. The serum levels of IL-6 among groups of the present study were with highest level in group 2 with *Toxoplasma* while the level of IL-6 in group 3 was with lower level compared with other groups.

Concentration of TGF-β1 in the study groups: The value for TGF-β1 in group1 was (2176.30±474.74) pg/ml, in group 2 was (1710.63±149.56) pg/ml and in group 3 was (1099.76±289.01) pg/ml as shows in the table 4. The highest level found in group 1 and the lowest level in group 3 and there was significant difference at (p<0.001) between different groups

Table 1: Distribution of studied groups according-to TORCH by rapid test and by ELISA test.

Study groups	TORCH by rapid test	TORCH by ELISA test
Recurrent abortion with sero-negative for TORCH test (group1)	39(18.6%)	36(17.1%)
Recurrent abortion with sero+ve for anti- <i>Toxoplasma</i> antibody (group2)	50(23.8%)	52(24.8%)
Pregnant normal without any abortion as a control group (group3)	30(14.3%)	30(14.3%)
Recurrent abortion with different cause (group4)	91(43.3%)	92(43.8%)
Total	210(100%)	210(100%)

Table (2): Seroprevalence of IgM and IgG among study groups according-to TORCH of ELISA test.

Pathogens	IgM %	IgG %	(IgM&IgG) %	Total
Toxoplasma gondii	18 (8.5%)	30 (14.3%)	4(1.9%)	52
Cytomegalovirus	17 (8.09%)	28 (13.3%)	5(2.4%)	50
Rubella	9 (4.3%)	15 (7.1%)	6(2.9%)	30
Herpes Simplex Virus	3 (1.5%)	8 (3.8%)	1(0.4%)	12
Healthy pregnant	0 (0%)	0 (0%)	0 (0%)	0
Total	47 (22.3%)	81(38.5%)	16(7.6%)	144

Concentration of IL-6 in the study groups.

Table 3: Serum level of IL-6 in the study groups.

Study groups	Number	Mean \pm SD (pg/ml)
Recurrent abortion with sero-negative for TORCH test (group1)	30	474.50 \pm 158.86 *
Recurrent abortion with sero-positive for anti- <i>Toxoplasma</i> antibody (group2)	30	629.60 \pm 115.45 *
Pregnant normal without any abortion as a control group (group3)	30	247.03 \pm 46.00 *
Total	90	---
LSD value	---	85.5
P-value	---	0.001

*= mean significant difference at ($p \leq 0.001$) by a nova.

Table 4: Serum level for TGF- β 1 in difference groups.

Study groups	Number	Mean \pm SD (pg/ml)
Recurrent abortion with sero-negative for TORCH test (group1)	30	2176.30 \pm 474.74 *
Recurrent abortion with sero-positive for anti- <i>Toxoplasma</i> antibody (group2)	30	1710.63 \pm 149.56 *
Pregnant normal without any abortion as a control group (group3)	30	1099.76 \pm 289.0 *
Total	90	---
LSD value	---	244.19
P-value	---	0.001

*= mean significant difference at ($p \leq 0.001$).

the rate expanded with age from the (15-19) years the rate was (5.6%) and from (40-44) years the rate was (16.7%)¹⁷.

The National Health and Nutrition Examination Survey (NHANES), found that the prevalence of toxoplasmosis has declined in the past decade¹⁸. Also, Acharya in 2014 showed that in Nepal the women with spontaneous abortions had IgG for *T. gondii* in rate (77.9%)¹⁹. Other Study was done in Egypt on pregnant women found the prevalence of *T. gondii* in the age (21-30) years was (50.8%)²⁰. Study done in Mexico on pregnant women infected with *T. gondii* recorded that IgG and IgM antibodies level in rate (6.1%), (9.5%) respectively²¹. Result was recorded by Hadi in 2016 in Iraq showed that in aborted women the percentage rate for IgG was (35.4 %) and for IgM was (3.2 %) ²². Also, in Iraq the women with spontaneous abortions had IgG and IgM for *T. gondii* in rate (31.5%), (7.6%) respectively²³. The percentage of IgG and IgM for *T. gondii* in aborted women its (22.8%) and (35.1%) respectively²⁴.

The role of some infection in recurrent abortion has been intensely investigated during the past decades by viruses, especially CMV have been more consideration since they can produce chronic/recurrent intrauterine infections. CMV causes both primary and recurrent (reactivation or

DISCUSSION

Seroprevalence of IgM and IgG among study groups: The rate of IgM antibody refers to acute infection, while IgG antibodies may increase in the 2-4 weeks after infection and gradually rises for many weeks then remain in low level for the rest time, while repeated exposure to the infection may cause high antibody rate for longer time¹².

Toxoplasma, *herpes*, *Rubella* and *Cytomegalovirus* are known to cause infection in uterus often responsible for abortion, still birth, premature delivery and congenital malformation, detection and treatment of such infections can prevent morbidity and mortality of the infants born to such mothers¹³.

The seroprevalence of *T. gondii* infections ranges between 7.7 and 76.7% in different countries like in United Kingdom (7.7- 9.1%); Norway (10.9%); India, (45%); Brazil(50-76%) and Nigeria (75.4%)¹⁴.

Turbadkar in 2003 revealed that in pregnant women which have history of bad pregnancies the percentage of IgG *T. gondii* antibody was (42.10%) and IgM was (10.5%)¹⁵. Study was done in India the rate of toxoplasmosis in pregnant women for IgG was (45%) and only seven women were found to have IgM in rate (3.3%)¹⁶. Nash (2005) reported that in United Kingdom the average of *Toxoplasma* in pregnant women for IgG was (9.1%), also obviously,

positive for HSV IgG antibodies was (28.9%), and for IgM antibodies was (2.2%)⁴³, Hasan in 2013 found that in pregnant women the rate of *herpes simplex virus* of IgG and IgM antibody were (2.19%), (2.19%) respectively⁴⁴. Study done in Nepal on the women with spontaneous abortions found that the HSV IgG antibody in rate (36.4%)¹⁹. While in Nigeria the titer of HSV IgM in pregnant women was (2.8%)⁴⁵. Other study in India showed that in aborted women the titer of HSV IgM was (30.10%)⁴⁶.

Differences between studies results can be due to the several laboratories utilize different techniques used in these aspects, type of test that had been used and the number or types of sample tested for infection and it is well-known that epidemiology of infection is different among different populations. Also, patients' characteristics have been different among studies.

Measurement of cytokines levels in the serum:

Pregnancy requires physiological adaptations in all maternal systems, including the immune system⁴⁷. Cytokines, as critical immunoregulatory molecules, responsible for determining the nature of an immune response, have been shown to influence on all steps of reproduction and playing a fundamental role in pregnancy outcome⁴⁸.

Interleukin-6 is an early and sensitive marker of inflammation, the major function of it was the contribution to immune response through the action of lymphocytes and consider a mediator responsible for producing acute phase proteins and increased cytotoxic activity of NK cells⁴⁹.

In this study, group 1 appeared high serum level of IL-6 compared with group 3, that indicate peripheral blood lymphocytes of women with recurrent spontaneous abortion secrete high level of IL-6 may explain the role of this cytokines in the pathogenicity of recurrent spontaneous abortion⁵⁰. Study done by Bakir in 2010 found that IL-6 increased in recurrent spontaneous abortion more than in healthy pregnant⁵¹.

Also, Hua in 2013 recorded that in the rat the concentration of IL-6 was significantly higher in recurrent miscarriage than normal pregnant⁵². On the other hand, Makhseed in 2000 showed that the serum level of IL-6 was significantly higher in pregnant women than in spontaneous abortion⁵³. Also, Koumantaki in 2001 recorded that the reduced plasma levels of IL-6 in women with spontaneous abortion may be related to the underlying etiopathogenetic mechanisms⁵⁴. Other study done by Ahmed in 2008 demonstrated that IL-6 level was lower in women with RSA than in those undergoing normal delivery⁵⁵. In sporadic miscarriage found that the increased in plasma levels of (IL-6)⁵⁶.

reinfection), viral strain variation may contribute to reinfection, and low maternal IgG avidity may unexpectedly promote transmission of virus across placenta and cause production of toxic metabolites, fetal loss, placental disorder, and chronic endometrial infection^{25,26}. Uyar in 2008 reported that in Turkey the pregnant women infected with CMV were had IgG and IgM in rate (97.3%) and (1.0%) respectively²⁷. Other study done in Sudan revealed that the prevalence of CMV IgG in pregnant women was 97.5%, while IgM was (6%)²⁸. Sherkat in 2014 recorded that in Iran CMV was significantly higher in recurrent pregnant loss (RPL) than the women without history of abortion and the titer of CMV IgG was (90.6%)²⁹. Study in Iraq on women with bad obstetric history (BOH) found that IgM and IgG antibodies to *Cytomegalovirus* in rate (7.2%), (96.6%) respectively³⁰. Other study in India, also on women with (BOH) the percentage of infection with CMV was (14.6%)³¹. Al-Baiati in 2014 showed that in aborted women the percentages of CMV for both IgG and IgM were (85%), (10%) respectively³². Also, other study in Iraq found that 90 pregnant women with an average age of 23 years had CMV IgG in rate (98.9%), while CMV IgM was (1.1%)³³.

Rubella is caused by RNA virus of paramyxovirus group. The pregnant women if contact with *Rubella* during the first 20 weeks of pregnancy will be harmfully affected by the virus. Its readily attacks the placenta and fetus then lead to miscarriage or stillbirth³⁴. Study was done in Iraq in aborted women with *Rubella* antibodies was positive for IgG in rate of thirty-four percent³⁵. Other study in Turkey the seropositive of the pregnant women for *Rubella* IgG, IgM were (96.1%) and (0.2%) respectively³⁶. Sadik in 2012 observed in India the seropositive of IgG for *Rubella* in pregnant women was (29.06%)³⁷. Acharya in 2014 showed that in Nepal the women with spontaneous abortions had *Rubella* IgG in rate (11.7%)¹⁹. Khudhair and Ahmed in 2015 recorded that the women in Iraq who had a one abortion the titer of *Rubella* IgG was (40%)³⁸.

Other report in Iraq revealed that in pregnant women with previous abortion the rate of *rubella* IgG was (57.8%) and *rubella* IgM was (26.6%), while the rate of *rubella* IgM in pregnant women without history of abortion was (3.8%)³⁹. Lamichhane in 2016 found that in spontaneous miscarriage the percentage rate for IgG was (43.68%) and for IgM was (7.76%)⁴⁰.

Also, pregnant women infected with *herpes simplex virus* can result neonatal, premature labor and abortion⁴¹. Biswas in 2011 reported that the pregnant woman in Indian the rate of IgG antibodies for HSV was (8.7%)⁴². Also, some studies in Iraq like Al-Marzoqi in 2012 recorded in pregnant women the

dosages of TGF- β 1, TGF- β 2, TGF- β 3 and its receptors, have been found in placentas after miscarriage, often with conflicting findings, highlighted by either decreased expression^{76,77} or unmodified protein levels⁷². The higher level of TGF- β 1 in pregnant women with miscarriage, compared with healthy pregnant women⁷⁸.

In this study, TGF- β 1 was higher in group 2 compared with group3. Also, previous studies showed that *T. gondii* induced TGF- β secretion by immune cells as an anti-inflammatory agent to decrease the autoimmune responses and the serum levels of TGF- β increased at an early phase of infection with *Toxoplasma*⁷⁹. Another investigation also demonstrated that *T. gondii* induced macrophages apoptosis through autocrine TGF- β signaling⁸⁰.

CONCLUSION

The current study of IL-6 serum levels were highest in women with recurrent abortion who had positive Anti-Toxoplasma Ab as compared with other groups while TGF- β 1 serum levels were highest among recurrent abortion women with undefined causes as compared with other groups.

REFERENCE

1. Williams and Wilkins. The Johns Hopkins Manual of Lippincott Gynecology and Obstetrics, (4thed.). 2012, p 438.
2. Tang A. W. and Quenby S. Recent thoughts on management and prevention of recurrent early pregnancy loss. *Current Opinion in Obstetric Gynecology*. 2010, 22(6): 446–451.
3. Abdul-Hadi CHabuk F. R. H., Al-Saadi H. K. Z. and Al-Hamairy A. K. Effect of the Experimental Infection with *Toxoplasma gondii* on some Biochemical aspects and Histological Changes for the Liver and Spleen in Female Rats. *International Journal of PharmTech Research*. 2016, 5(11): 142-150.
4. Malarvizhi A., Viswanathan T., Lavanya V., Sheeba Malar S. A. and Moorthy K. Seroprevalence of *Toxoplasma gondii* in pregnant women. *Public Health and Epidemiology*. 2012, 4(6):170-177.
5. Raghupathy R. The immunology of unexplained recurrent spontaneous abortion: cytokines as key mediators. *Bulletin of the Kuwait Institute for Medical Specialization*. 2003, 2(1):32-38.
6. Piccinni M. R., Lombardelli L., Logiodice F., T helper cell mediated-tolerance towards fetal allograft in successful pregnancy. *Clinical and Molecular Allergy*. 2015, 13(1): 9-12.
7. Nair R., Khanna A. and Singh K. Association of Interleukin 1 beta (IL-1B) gene polymorphism with early pregnancy loss risk in the North Indian population. *International Journal of Immunogenetics*. 2013,41(1):443 -450.
8. Zenclussen A. C., Blois S., Stumpo R., Olmos S., Arias K., Malan Borel I., Roux M. E. and Margni R. A. Murine abortion is associated with enhanced interleukin-6 levels at the fetomaternal interface. *Cytokine*. 2003, 24(4):150-160.
9. Dkhil A. S., Hussain F. and Muttar A. A. Association Between Interleukin-6(IL-6) and Thrombocytosis in Rheumatoid Arthritis Patients. *International Journal of PharmTech Research*. 2016, 9(4): 307-312.

While Rageb in 2015 showed that the IL-6 level was higher in *toxoplasma* group than control group⁵⁷, this may be critical role of IL-6 to resistancethe infection with *T. gondii*⁵⁸ and contributing tottransformationfrom acute to chronic inflammation⁵⁹.

El-Hashimi in 2014 showed that IL-6 in aborted women infected withtoxoplasmosis was significantly higher compared to aborted women without toxoplasmosis⁶⁰. Hafedh in 2015 recorded that in aborted women the concentration of IL-6 was more than in healthy womenbecause IL-6 play a vital role during the infection with *T. gondii* in aborted women⁶¹. Previous studies showed that the high level of IL-6 in the patients with toxoplasmosis may explain the presence of anti- *T. gondii* antibody in serum of patient, this may occur early in infection before the immune response modified from Th2 into Th1 that responsible for the several cases of abortion^{49,62}.

Tumor growth factor- β eta may contribute to the regulation of maternal immune responses against the fetal allograft, and thereby prevent immunological rejection of the fetus⁶³. TGF- β 1 is secreted by many cell types, including peripheral blood mononuclear cells and T regulatory lymphocytes⁶⁴.

Tumor growth factor- β eta1, exerting a predominantly anti-inflammatory effect and regulate several aspects of pregnancy, thereby contributes to maintaining immune tolerance^{65,66,67}. The plasma level of TGF- β 1 in recurrent spontaneous abortion was significantly higher compared with pregnant normal. Also, explained the increased in the production this molecule may be a risk factor for the pregnancy outcome⁶⁸.

Several studies suggested that TGF- β 1 may be involved in reproductive related disorders, such as preeclampsia and recurrent spontaneous abortion, although data were controversial^{69,70}. An imbalance between the effector and regulator cells would lead to reproductive failure and related pregnancy disorders⁷¹.

Other studies found that the levels of TGF- β 1, TGF- β 2 and TGF- β 3 were unaltered in the plasma of spontaneous abortion⁷². TGF- β 1 has both endocrine and paracrine actions, so in situ placental expression are more relevant than analysis of plasma levels⁶⁷. Gutcher in 2011 reported that TGF β 1 was highly expressed by Th17 cells and acted in a predominantly autocrine manner to maintain Th17 cells *in vivo*, the role of TGF β for activated T cell-produced in promoting the differentiation of Th17 cells and controlling inflammatory diseases⁷³. The capacity of TGF- β 1 to regulate the cytokine network that controls trophoblastgrowth&uterine invasion^{74,75}.

Other studies have examined TGF- β 1 expression in the placenta of RPL cases, reduced

27. Uyar Y., Balci A., Akcali A. and Cabar C. Prevalence of rubella and cytomegalovirus antibodies among pregnant women in northern Turkey. *New Microbiological*. 2008, 31(4):451-455.
28. Khairi S. I., Intisar K. S., Enan K. H., Ishag M. Y., Seroprevalence of cytomegalovirus infection among pregnant women at Omdurman Maternity Hospital, Sudan. *Journal of Medical Laboratory and Diagnosis*. 2013, 4(4):45-49.
29. Sherkat R., Meidani M., Zarabian H., Rezaei A. and Gholamrezaei A. Seropositivity of cytomegalovirus in patients with recurrent pregnancy loss. *Journal Research of Medical Science*. 2014, 19(1): S22-S25.
30. Aljumaili Z. K. M. and Alsamarai A. M. Risk factors for bad obstetric history in Kirkuk women, Iraq. *International Journal of Infection Microbiology*. 2013, 2(3): 70-77.
31. Faldu B. R., Panchal P., Patel H. L. and Patel A. Study of Seropositivity for TORCH Infections in Women with Bad Obstetric History. *International Journal of Advanced Research*. 2015, 3(8):1324 -1332.
32. Al-Baiati A. M. H. P., Muhsin M. A. and Jabbar R. N. Seroprevalence of Human Cytomegalovirus (HCMV) in aborted women in Baghdad. *International Journal of Current Microbiology and Applied Sciences*. 2014, 3(2): 97-102.
33. Altayeb M. A., Mokhtar S. I., Adam M. E., Mohammed S I and Musa H H. Detection of primary CMV infection in Sudanese pregnant women by IgG avidity test. *Asian Pacific Journal of Tropical Disease*. 2016, 6(10): 816-818.
34. Kolawole O. M., Anjorin E. O., Adekanle D. A., Kolawole C. F., and Durowade K. A. Seroprevalence of Rubella IgG Antibody in Pregnant Women in Osogbo, Nigeria. *International Journal of Preventive Medicine*. 2014, 5(3): 287.
35. Abdul-Karim E. T., Abdul-Muhyen N. and Al-Saadie M. *Chlamydia trachomatis* and *rubella* antibodies in women with full-term deliveries and women with abortion in Baghdad. *Eastern Mediterranean Health Journal*. 2009, 15(6):1407.
36. Tamer J. S., Dundar D. and Caliskan E. Seroprevalence of *Toxoplasma gondii*, rubella and cytomegalovirus among pregnant women in western region of Turkey. *Clinical and Investigative Medicine*. 2009, 32 (1): E43-E47.
37. Sadik M. S., Fatima H., Jamil K. and Patil C. Study of TORCH profile in patients with bad obstetric history. *Biology and Medicine*. 2012, 4 (2): 95-101.
38. Khudhair M. K. and Ahmed R. I. Effect of anti-Rubella (IgG) on a number of abortion in pregnant women of living in Diyala Province. *Journal of Dental and Medical Sciences*. 2015, 14(5):60-63.
39. Mohammed L. M. Prevalence of rubella virus in pregnant women in Kirkuk city-Iraq. *Kirkuk University Journal /Scientific Studies*. 2015, 10(1): 47-57.
40. Iamichhane S., Subedi S., Pokhare S., Chetri M. and Banerjee B. Relationship of Torch Profile in First Trimester Spontaneous Miscarriage. *Journal of Nobel Medical College Available*. 2016, 5(2):17-21.
41. Kimberlin D. W. Herpes simplex virus infections of the newborn. *Seminar Perinatology*. 2011, 31(1):19-25.
42. Biswas D., borkakoty B., Mahanta J., walia K., Saikia L., Akoijam B. S., Jampa L., Kharkongar A. and Zomawia E. Seroprevalence and risk factors of herpes simplex type-2 infection among pregnant women in northeast India. *BioMed Central Infectious Diseases*. 2011, 11(3):325-330.
43. Al-Marzoqi A. H. M., Kadhim R. A., Al-Janabi D. K. F., Hussein H. J. and Al-Tae Z. M. Seroprevalence study of IgG and IgM antibodies to *toxoplasma*, *rubella*, *cytomegalovirus*, *Chlamydia trachomatis* and *herpes simplex II* in pregnancy women in Babylon province. *Journal of Biology Agriculture and Healthcare*. 2012, 2(10):159-64.
44. Hasan A. S. H., Al-Duliami A. A., Hwaid A. H., Mageed W. A. and Fadeel Z. G. Seroprevalence of Anti- Herpes Simplex Virus Type2 IgG, IgM Antibodies Among Pregnant Women in Diyala Province. *Diyala Journal of Medicine*. 2013, 5(1):36.
10. Mohammed A. and Hassan A. J. Estimation of pro and anti-inflammatory cytokines related with obese individuals in Babylon/Iraq. *International Journal of ChemTech Research*. 2016, 9(12):482-488.
11. Korn T., Bettelli E., Oukka M., Kuchroo V. K. IL-17 and Th17 Cells. *Annual Review of Immunology*. 2009, 27(4):485-517.
12. Kumar A., Seringhaus M., Sarnovsky R. J., Biery M. C., Umansky L., Piccirillo S., Heidtman M., Cheung K. H., Dobry G. J., Gerstein M. B., Craig N L and Snyder M. Large-Scale Mutagenesis of the Yeast Genome Using a Tn7-Derived Multipurpose Transposon. *Genome Research*. 2004, 14(10A):1975-1986.
13. Agrawal A., Singh S. K., Maheshwari R. K. and Goya J. K. Study of Serological Profile of Torch Infections in Women with Bad Obstetric History. *Microbiology*. 2016, 6 (5):2249-555X.
14. Gulden S. T., Devrim D. and Eray C. Seroprevalence of *Toxoplasma gondii*, *Rubella* and *Cytomegalovirus* among pregnant women in western region of Turkey. *Clinical Investigative Medicine*, 2009. 32 (1): 43-47.
15. Turbadkar D., Mathur M. and Rele M. Seroprevalence of torch infection in bad obstetric history. *Indian Journal of Medicine Microbiology*. 2003, 21(2): 108-110.
16. Singh S. and Pandit A. J. Incidence and prevalence of toxoplasmosis in Indian pregnant women: a prospective study. *American Journal of Reproductive Immunology*. 2004, 52(3):276-283.
17. Nash J. Q., Chissel S., Jones J., Warburton F. and Verlander N. Q. Risk factors for toxoplasmosis in pregnant women in Kent, United Kingdom, *Epidemiological and Infection*. 2005, 133(3):475-483.
18. Jones J. L., Kruszon-Moran D., Sanders-Lewis K. and Wilson M. *Toxoplasma gondii* infection in the United States, 1999-2004, decline from the prior decade. *The America Journal of Tropical Medicine and Hygiene*. 2007, 77(3):405-410.
19. Acharya D., Shrestha A., Bogati B., Khanal K., Shrestha S. and Gyawali P. Serological screening of TORCH agents as an etiology of spontaneous abortion in Dhulikhel hospital, Nepal. *American Journal of Biomedical and Life Sciences*. 2014, 2(2): 34-39.
20. Kamal A. M., Ahmed A. K., Abdellatif M. Z. M., Tawfik M., and Hassan E. E. Seropositivity of Toxoplasmosis in Pregnant Women by ELISA at Minia University Hospital, Egypt. *Korean Journal of Parasitology*. 2015, 53(5): 605-610.
21. Alvarado-Esquivel C., Sifuentes-Alvarez A., Narro-Duarte S. G., Estrada-Martínez S., Díaz-García J. H., Liesenfeld O., Martínez-García S. A. and Canales-Molina A. Seroepidemiology of *Toxoplasma gondii* infection in pregnant women in a public hospital in northern Mexico. *BMC Infectious Diseases*. 2006, 6(2):113-118.
22. Hadi H. S., Kadhim R. A. and Al-Mammori R. T. O. Seroepidemiological aspects for *Toxoplasma gondii* infection in women of Qadisiyah province. Iraq *International Journal of PharmTech Research*. 2016, 9(11): 252-259.
23. Al-Kalaby R. F., Sultan B. A. and AL-Fatlawi S. N. Relationship between *Toxoplasma gondii* and abortion in aborted women in Najaf province. *Journal of Kerbala University*. 2016, 14(1):98-104.
24. Abbas S. S. and Al-Hamairy A. K. Molecular Study of *Toxoplasmosis* and Its Relationship With Some Parameters (TSP, Globulin and Albumin) Among pregnant and aborted women in the Babylon Province Iraq. *International Journal of PharmTech Research*. 2016, 9(9): 366-380.
25. Nigro G. and Adler S P. Cytomegalovirus infections during pregnancy. *Current Opinion in Obstetric Gynecology*, 2011. 23(2):123-128.
26. Nigro G, Mazzocco M., Mattia E., Di Renzo G. C., Carta G. and Anceschi M. M. Role of the infections in recurrent spontaneous abortion. *Journal Maternal Fetal Neonatal Medicine*. 2011, 24(8):983-989.

63. Power L. L., Popplewell E. J., Holloway J. A., Diaper N. D., Warner J. O. and Jones C. A. Immunoregulatory molecules during pregnancy and at birth. *Journal Reproductive Immunology*. 2002, 56:19–28.
64. Li M. O., Wan Y. Y. and Flavell R. A. T cell-produced transforming growth factor-beta1 controls T cell tolerance and regulates Th1 and Th17-cell differentiation. *Immunity*. 2007, 26(5):579–591.
65. Li M. O., Sanjabi S., Flavell R. A. Transforming growth factor-beta controls development, homeostasis, and tolerance of T cells by regulatory T cell-dependent and -independent mechanisms. *Immunity*. 2006, 25(3):455-471.
66. Taylor A.W. Review of the activation of TGF- β in immunity. *Journal Leukocyte Biology*. 2009, 85(1):29–33.
67. Ingman W. V. and Robertson S. A. The essential roles of TGF β 1 in reproduction. *Cytokine Growth Factor Review*. 2009, 20(3):233-239.
68. Amani D., Dehaghani A. S., Zolghadri J., Ravangard F., Lack of association between the TGF- β 1 gene polymorphisms and recurrent spontaneous abortion. *Journal of Reproductive Immunology*. 2005, 68(1-2): 91-103.
69. Benian A., Madazli R., Aksu F., Uzun H. and Aydin S. Plasma and placental levels of interleukin-10, transforming growth factor-beta1, and epithelial-cadherin in preeclampsia. *Obstetrics and Gynecology*. 2002, 100(2):327–331.
70. Huber A., Hefler L., Tempfer C., Zeisler H., Lebrecht A. and Husslein P. Transforming growth factor-beta1 serum levels in pregnancy and preeclampsia. *Acta Obstetric Gynecology Scandinavia*. 2002, 81(2):168–171.
71. Lee S. K., Kim J. Y., Lee M., Gilman-Sachs AKwak-Kim J. Th17 and Regulatory T cells in Women with Recurrent Pregnancy Loss. *American Journal of Reproductive Immunology*. 2012, 67(4): 311–318.
72. Ball E., Robson S. C., Aiyis S., Lyall F. and Bulmer J. N. Expression of TGF beta in the placental bed is not altered in sporadic miscarriage. *Placenta*. 2007, 28(8-9):965-971.
73. Gutcher I., Donkor M. K., Ma Q., Rudensky A.Y., Flavell R. A.. Autocrine Transforming Growth Factor- β 1 Promotes in vivo Th17 Cell Differentiation. *Immunity*. 2011, 34(3): 396.
74. Tse W. K., Whitley G. S. and Cartwright J. E. Transforming growth factor-beta1 regulates hepatocyte growth induced trophoblast motility and invasion. *Placenta*. 2002, 23(10):699.
75. Lash G. E., Otun H. A., Innes B. A., Percival K., Searle R. F., Regulation of extravillous trophoblast invasion by uterine natural killer cells is dependent on gestational age. *Human Reproductive*. 2010, 25(5):1137-1145.
76. Cheng L. H. and Cao Y. X. Study on the correlation of transforming growth factor beta1 and its receptors with spontaneous abortion after in vitro fertilization and embryo transfer. *Zhonghua Fu Chan KeZaZhi*. 2005, 40(5):299-301.
77. Giannubilo S. R., Landi B., Pozzi V., Sartini D., The involvement of inflammatory cytokines in the pathogenesis of recurrent miscarriage. *Cytokine*. 2012, 58(1):50–56.
78. Magdoud K., Herbepin V. G., Granados V., Messaoudi S., Hizem S., Bouafia N., Genetic variation in TGF β 1 gene and risk of idiopathic recurrent pregnancy loss. *Molecular Human Reproductive*. 2013, 19(7):438-443.
79. Wagner A., Förster-Waldl E., Garner-Spitzer E., Schabussova I., Kundi M., Pollak A., Scheiner O., Joachim A. and Wiedermann U. Immunoregulation by *Toxoplasma gondii* infection prevents allergic immune responses in mice. *International Journal of Parasitology*. 2009, 39(4):465-72.
80. D'Angelillo A., De Luna E., Romano S., Bisogni R., Buffolano W., Gargano N., Del Porto P., Del Vecchio L., Petersen E. and Romano M. F. *Toxoplasma gondii* Dense Granule Antigen 1 stimulates apoptosis of monocytes through autocrine TGF- β signaling. *Apoptosis*. 2011, 16(6):551-562.
45. Okonko I. O., Cookey T., Okerentugba P. O. and Frank-Peterside N. Serum HSV-1 and -2 IgM in pregnant women in Port Harcourt, Nigeria. *Journal of Immunoassay Immunochemistry*. 2015, 36(4):343-358.
46. Tiwari S., Arora B. S. and Diwan R. TORCH IgMseroprevalence in women with abortions as adverse reproductive outcome in current pregnancy. *International Journal of Research Medical Sciences*. 2016, 4(3): 784-788.
47. Luppi P. How immune mechanisms are affected by pregnancy. *Vaccine*. 2003. 21(24):3352-3357.
48. Sharief M., Mohammed R. and Shani W. S. The role of Th1 and Th2 cytokines among women with recurrent spontaneous miscarriage. *Scientific J Med Sci*. 2014, 3(7) 345-351.
49. Matowicka-Karna J, Dymicka-Piekarska V. and Kemona H. Does *Toxoplasma gondii* infection affect the levels of IgE and cytokines (IL-5, IL-6, IL-10, IL-12, and TNF-alpha)? *Clinical and Developmental Immunology*. 2009, 2009(374696):4-9.
50. Prins J. R., Gomez-Lopez N. and Robertson S. A. Interleukin-6 in pregnancy and gestational disorders. *Journal of reproductive immunology*. 2012, 95 (1-2):1-14.
51. Bakir W. A., Zynab S., Abdul-gany A. The Role of IL-6, IL-10 and IFN- γ mRNA in Women with Recurrent Abortion. *Iraqi Journal of Cancer and Medical Genetics*. 2010, 3(1):55-63.
52. Hua F., Li C. H., Wang H. and Xu H. G. Relationship with expression of COX-2, TNF-B, IL-6 and autoimmune-type recurrent miscarriage. *Asian Pacific Journal of Tropical Medicine*. 2013, 6(12): 990-994.
53. Makhseed M., Raghupathy R., Azizieh F., Farhat R., Hassan N. and Bandar A. Circulating cytokines and CD30 in normal human pregnancy and recurrent spontaneous abortions. *Human Reproductive*. 2000, 15 (9): 2011-2017.
54. Koumantaki Y., Matalliotakis I., Sifakis S., Kyriakou D., Neonaki M., Detection of interleukin-6, interleukin-8, and interleukin-11 in plasma from women with spontaneous abortion. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*. 2001, 98(1):66-71.
55. Ahmed D. W. Effects of Interleukin-2 (IL-2) and Interleukin-6 (IL-6) in Recurrent Spontaneous Abortion (RSA). *Iraqi Journal of Pharmaceutical Sciences*. 2008, 17(2):74-79.
56. Calleja-Agius J., Jauniaux E., Pizzey A. R and Muttukrishna S. Investigation of systemic inflammatory response in first trimester pregnancy failure. *Human Reproductive*. 2011, 27(2): 349-357.
57. Rageb O., Abd El-Maksoud H. A., Afaf D. A. and Fathy K. A. Biochemical Effect of *Toxoplasma* Infestation on Immunity and Inflammatory Markers In Aborted Women. *Benha veterinary medical journal*. 2015, 28(1): 125-132.
58. Passos S. T., Silver J. S., O'Hara A. C., Sehy D., Stumhofer J. S. IL-6 promotes NK cell production of IL-17 during toxoplasmosis. *Journal of Immunology*, 2010. 184(4): 1776.
59. Trunov A., Obukhova O., Gorbenko O., Shvayk A. and Trunova L. Cytokines and Infertility Influence of Cytokines and Local Inflammation in Women of Reproductive Age with Infertility. *Journal of Cytokine Biology*. 2016, 1(1):102-109.
60. El-Hashimi W. K., Khalila H. I., Al-Mayahb Q. S. The role of cytokines, TNF- α , IL-6 and pregnancy associated hormones in *Toxoplasma gondii* induced abortion. *European Journal of Experimental Biology*. 2014, 4(6):98-104.
61. Hafehd A. A. Detection of IL-6 Level and the Diagnostic Agnostic Value of Anti- *Toxoplasma Gondii* IgM and IgG in Aborted Women in AL-Nasiriya City/IRAQ. *European Journal of Biomedical and Pharmaceutical Sciences*. 2015, 2(5): 621
62. Rezende-Oliveira K., Silva N. M., Mineo J. R. and Rodrigues-Junior V. Cytokines and chemokines production by mononuclear cells from parturient women after stimulation with live *Toxoplasma gondii*. *Placenta*. 2012, 33(9):682–687.